# Metagenomic Fingerprinting of Rainwater Harvesting Systems

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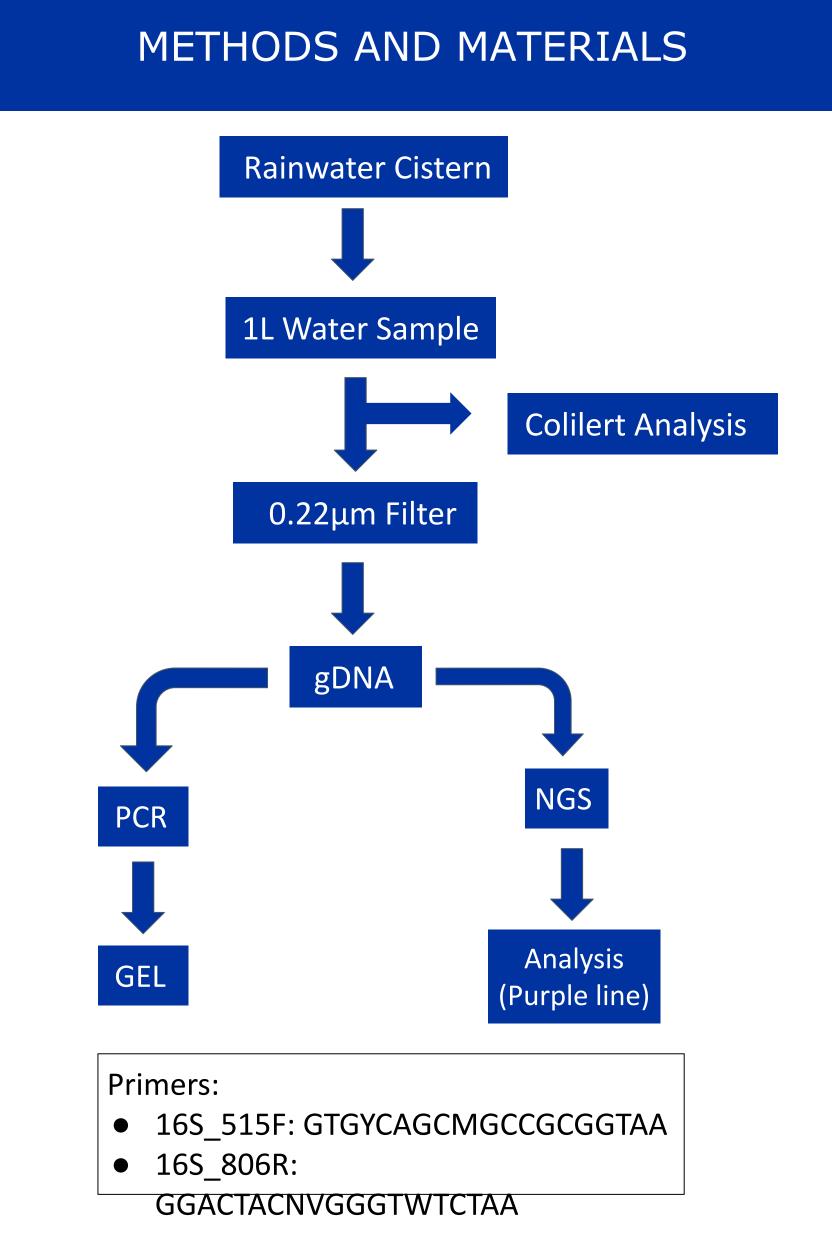
# BACKGROUND

- Rainwater collection systems are used to collect and store rainwater from the environment (figure 1).
- These <u>collection systems</u> tend to <u>accumulates containments</u> like fecal matter, insect larvae, bacteria, and pathogens, which could lead to contamination and possible health <u>hazards</u>.
- Colilert<sup>®</sup> and 16s rRNA sequencing were used to detect bacteria present in systems.
- A Colilert<sup>®</sup> detection system uses MPN to estimate coliform bacteria populations present in water<sup>1,2</sup>.
- The 16s rRNA gene was used to identify bacteria using metagenomic sequencing.
- "Metagenomic Fingerprinting" provided an overview of the diversity of bacteria in the rainwater collection systems<sup>3</sup>.
- Metagenomic analysis has rarely been used on samples of rainwater collection systems; however, it is an ideal method of determining the microbial diversity in these systems.

# **RESEARCH QUESTION & HYPOTHESIS**

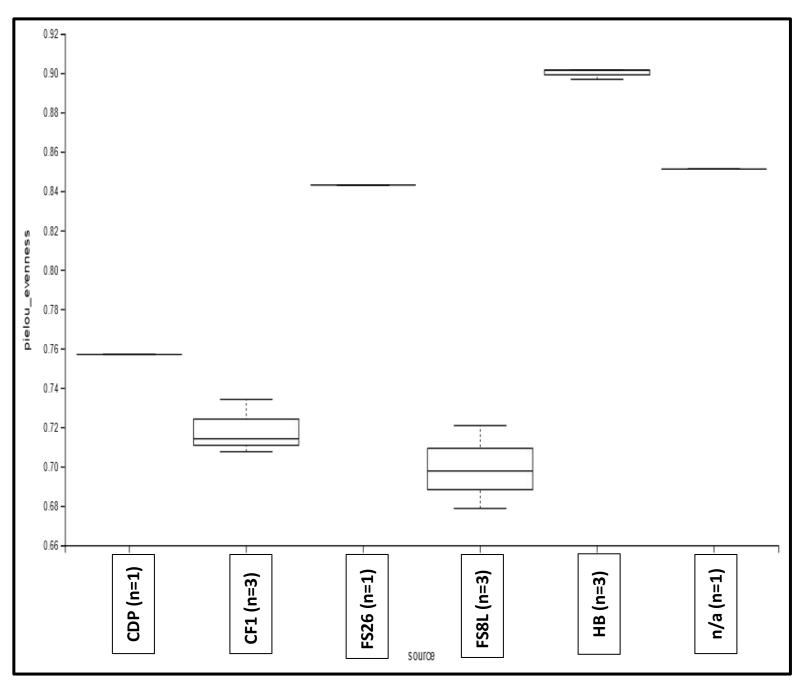
**Research Question:** What is the microbial diversity in rainwater harvesting systems and what factors affect that diversity?

**Hypothesis:** Application of metagenomic fingerprinting will demonstrate changes in the diversity of bacteria based on environmental factors.





(Source HB).



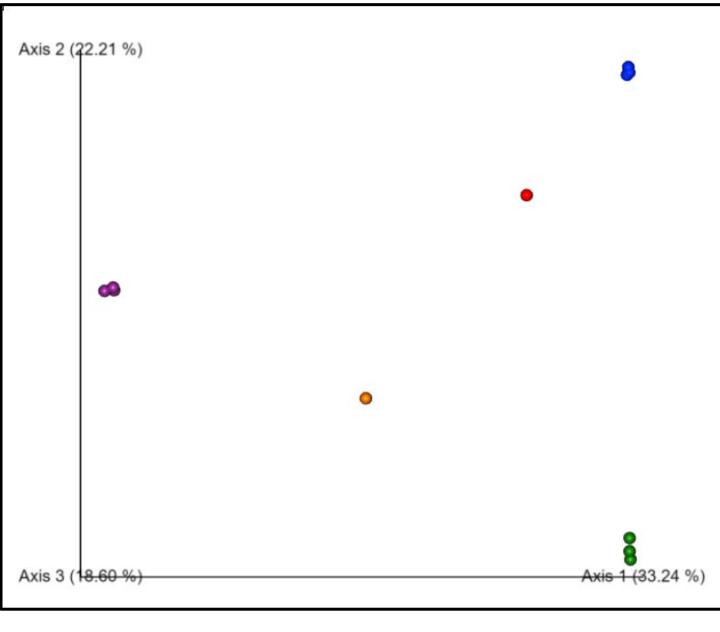


Figure 3. Beta Diversity using Bray Curtis **Distance.** (Purple: HB , Orange:FS26, Red: CDP, Blue:CF1, Green:FS8L)

Figure 1. Typical example of a **Rainwater Harvesting System** 



| Source<br>(Replicates of 3) | рН   | Tank Volume | Water Temperature (°<br>C) |  |
|-----------------------------|------|-------------|----------------------------|--|
| FS26                        | 5.93 | 5678        | 7.3                        |  |
| FS8L                        | 7.54 | 4542        | 9.7                        |  |
| НВ                          | 7.66 | 15141       | 6.9                        |  |
| CDP                         | 7.98 | 1155        | 5.2                        |  |
| CF1                         | 7.92 | 2082        | 13.3                       |  |

 Table 1. Metadata for water samples.

| Sample<br>Site | Average Coliform Concentration<br>(MPN/100ml) |  |
|----------------|---|--|
| FS8L           | >2419.6                                       |  |
| HB             | 19.67   |  |
| FS26           | 25.27   |  |

 
 Table 2. Average coliform presence by
 **Colilert**<sup>®</sup>. Only sites shown had coliform present. Remaining samples detected no coliform. Samples were performed in triplicates.

### Figure 2. Alpha Diversity using Pielou's Evenness. Alpha diversity is highly variable between sources.

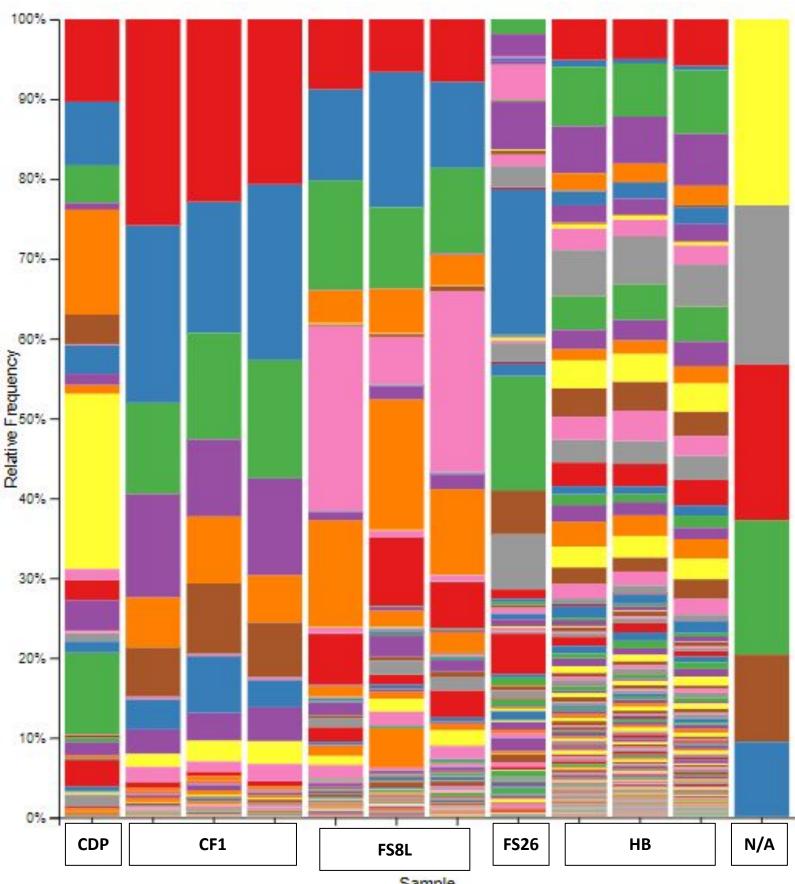


Figure 4. Taxonomic Diversity. Bar length indicates relative frequency of bacteria present.

|               | OTU (Operational<br>Taxonomic Unit) |                  | FS8L | НВ | CDP | CF1 | FS26 |
|---------------|-------------------------------------|------------------|------|----|-----|-----|------|
|               | Family Genus                        |                  |      |    |     |     |      |
|               | 1. Oxalobacteraceae                 | Polynucleobacter | Х    | _  | Х   | Х   | _    |
| Most Frequent | 2. Comamonadaceae                   | _                | Х    | х  | Х   | Х   | _    |
|               | 3. Chitinophagaceae                 | _                | Х    | х  | Х   | Х   | _    |
|               | 4. Rhodospirillaceae                | —                | -    | х  |     |     | Х    |
|               | 5. Sphingomonadaceae                | Novosphingobium  | Х    | _  | Х   | Х   | _    |
|               | 1. Legionellaceae                   | —                | Х    | х  | Х   | Х   | X    |
| Pathogenic    | 2. Enterobacteriaceae               | —                | х    | _  | _   | _   | _    |
|               | 3. Clostridiaceae                   | Clostridium      | х    | х  | _   | _   | _    |
|               | 4. Bacillaceae                      | Bacillus         | Х    | х  | _   | _   | _    |

Table 3. Presence or absence of OTU's in each source organized by frequency and recognized pathogens.



### CONCLUSIONS

- Colilert<sup>®</sup> detected the presence of coliform bacteria in some barrels
- Metagenomics Fingerprinting provides a robust indication of microbial diversity in rainwater sources
- Results show differences between each sources
- Alpha diversity indicates diversity within sources
- Beta diversity indicates diversity between sources
- Taxonomic diversity indicates a large amount of diversity for each source
- Limitations
  - Using pre packaged version of QIIME limited analysis and background information
  - 300 bp sequences of 16s rRNA were insufficient to identify OTU's to species.
  - Preliminary data includes only one time point

### FUTURE DIRECTIONS

- This preliminary data will be used in another study using EMA qPCR to quantify bacteria.
- A second analysis of a late summer sample will be conducted and compared for seasonal variability.
- We plan to continue sampling so we can improve statistical analysis.

### ACKNOWLEDGMENTS

- The Longwood Office of Student Research pr funding.
- Dr. Ray Enke, of James Madison University, c the metagenomic sequencing.
- Metagenomic Fingerprint analysis was comp **Cyverse DNA Subway**

### REFERENCES

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